
PROTECTION OF FOOD FROM FUNGAL PATHOGENS A REVIEW

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ABSTRACT

KEYWORDS

Phytopathogen,
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The profitability, quality, and output volume of plant production is significantly influenced by plant fungal diseases. These phytopathogens are persistent in getting past plant defences, which leads to diseases and quality losses that cost the US economy billions of dollars every year. Farmers have employed fungicides to manage the damage caused by plant pathogenic fungus in order to combat the epidemic of fungal plant diseases. Researchers and growers are looking for alternate solutions because of drawbacks including resistance development and environmental damage linked to these drugs. Materials and Procedures Using the search terms "plant fungal pathogen," "plant extracts," and "phytopathogens," several databases were consulted to learn more about research on protecting plants against plant fungal diseases. The best extractants and bioassay methods are suggested for use. Results: Plant fungal diseases have previously been treated with biological agents in addition to conventional fungicides. There are numerous instances when plant extracts or chemicals derived from plants have been employed on a broad scale as commercial fungus deterrents in agricultural and horticultural settings. The fact that plant extracts typically include many antifungal compounds is a benefit of this strategy. Consequently, if various substances have an impact on various metabolic processes, the development of pathogen resistance may be reduced. Plants grown with the use of plant extracts may also be marketed as organic. Numerous studies on potent antibacterial substances found in plant extracts with a focus on applications in human health have been released. To create acceptable, affordable, efficient, and sustainable botanical solutions that can be utilised to combat the epidemic of plant fungal infections, more study is necessary. Conclusions: The benefits of concentrating on plant fungal infections should be considered by scientists who have solely concentrated on using plants to control human and animal fungal illnesses. This strategy is considerably simpler to assess the efficacy in greenhouse or field studies, which could boost the food security for farmers in rural areas and result in financial incentives. Extracts may still be valuable in the floriculture sector even if they are hazardous..

INTRODUCTION

The majority of infections that occur in horticultural and agricultural settings are caused by plant fungal pathogens (Agrios, 2009). Fungal diseases are effecting both creature animal and Humans some fungal diseases like aspergillus infection are similar to viral diseases but fungal diseases have treatment while viral disease don't have any treatment (Samad, Hamza, Muazzam, Ahmer, Tariq, Javaid, et al., 2022; Samad, Hamza, Muazzam, Ahmer, Tariq, Javaid, et al., 2022; Samad et al, 2021; Samad, Hamza, Muazzam, Ahmad, Ahmer, Tariq, et al., 2022a, 2022b; F. M. S. Muthanna & Samad, 2022, Ibrahim et al., 2022; Mohammed et al., 2022) while bacterial disease also have treatment. (Mohammed et al., 2022b; F. M. Muthanna et al., 2022).

To target any plant, the phytopathogens have collectively developed strategies and methods (Knogge, 1996) that allow them to forcefully access nutrients for growth and

development (Horbach et al., 2011). These infections are sexually and/or asexually reproducing (Gould, 2009), and they are capable of defeating plant immune defences (Thomma et al., 2011; Zvereva and Pooggin, 2012). This harms the health, homeostasis, physiology, and in some situations, the systemic health of the plant (Agrios, 2005). Plant fungal pathogens must be able to grow on the surface of an appropriate host for plant fungal diseases to manifest. The right circumstances are necessary for the plant fungal pathogen's spores to germinate. This includes a suitable host, low molecular mass nutrients, and adequate humidity from rain or dew (Osheroov and May, 2001). By utilising self-inhibitors to delay germination until favourable conditions are present, fungal spores can survive for many years (Chitarra et al., 2004). For the hyphae to enter the host when the conditions are right, plant fungal diseases create infection structures such the appressorium and the infection peg (Schafer, 1994). Waxy hosts can serve as a vehicle for pathogens like *Colletotrichum gloeosporioides* to infect their hosts and cause illnesses in avocados (Podila et al., 1993). Different tactics can be employed by plant fungal infections to attack and infect their host. Others enter their host through wounds and the stomata, while certain germs enter by mechanical pressure and chemical activity (Knogge, 1998). Plants have evolved defences against fungus-related infections. This is an excellent reason to look into if plants contain antifungal chemicals (Eloff and McGaw, 2014). The pathogen associated molecular patterns (PAMP), PAMP-triggered immunity (PTI) inducible defence, and effector triggered immunity (ETI) defence are plant host defence systems that the plant fungal pathogens need to avoid (Thomma et al., 2011; Zvereva and Pooggin, 2012). Successful pathogens that are able to circumvent plant PTI and ETI, particularly those that have developed their avirulent genes to circumvent plant host R-resistance genes (Stergiopoulos and de Wit, 2009), produce catastrophic plant illnesses that can spark epidemics (Dean et al., 2012). Not all plant fungal infections can infect a single host with illness. Some have a restricted range of hosts, while others have a wide range. A variety of plant fungal diseases may be able to live on some plants (Agrios, 2005). Plant fungi can spread to new areas where they have never existed through the wind, birds, people, insects, water, and contaminated plant parts (Agrios, 2005; Rossman, 2009). Although not all fungi infect plants, plant fungi diseases attack all types of plants (Knogge, 1996). Collectively, these fungi are to blame for 80% of plant illnesses (El Hussein et al., 2014). Nearly 100 000 plant diseases are caused by 8000 different fungus species (Agrios, 2005). Based on the mode of infection, plant fungal pathogens can generally be divided into biotrophs, necrotrophs, and hemibiotrophs. Infections are caused by biotrophs, which feed on live tissues and do not harm their hosts. Some pathogens enter the host through the appressorium and use feeding organs like the haustoria to obtain nourishment from the cells around them. The hosts for the biotrophs are restricted, such as rust fungi and powdery mildew fungi (De Silva et al., 2016). Necrotrophs kill the afflicted area by infecting the living host. This is so that necrotrophic fungi, which require dead tissues to complete their life cycles, can exist. To obliterate the plant cells, these pathogens continuously create hydrolytic enzymes and poisons. In order for the pathogen to infect a particular host, such as *Cochliobolus carbonum*, necrotrophs create two different types of poisons, called host specific toxins. Second, some infections, such as *Sclerotinia sclerotiorum*, *Alternaria brassicicola*, and *Botrytis cinerea*, can infect and kill unrelated plant species thanks to broad-spectrum toxins (Wen, 2013). The hemi-biotrophs, such as *Colletotrichum*, infect their hosts through similar mechanisms to biotrophs before killing them as necrotrophs (Agrios, 2005).

METHOD RESEARCH

The research method used in this study is a qualitative descriptive method. The type of data used in this study is qualitative data, which is categorized into two types, namely primary data and secondary data. Sources of data obtained through library research techniques (library study) which refers to sources available both online and offline such as: scientific journals, books and news sourced from trusted sources. These sources are collected based on discussion and discussion from one information to another. Data collection techniques used in this study were observation, interviews and research. This data is analyzed and then conclusions are drawn

RESULT AND DISCUSSION

A. Problems With Agricultural Production And Food Degradation Brought On By Plant Fungal Infections

Agriculture can produce sustainable plant products that can reduce hunger and poverty (Alexandratos and Bruinsma, 2012). The agricultural industry has suffered significant losses as a result of epidemics like the late blight diseases of potatoes, cereal rusts and smuts, ergot of rye and wheat, brown spot of rice, coffee rust, Sigatoka disease of bananas, chestnut blight, the downy and powdery mildews of grapes, wheat stem rust, and rubber leaf blight. Reduced plant quality and quantity caused by plant fungal infections can potentially be a concern to human health. This could result in forced food migration, altered country economic outlooks, increased political unpredictability, and forced people migration (Anderson et al., 2004; Ellis et al., 2008; Gould, 2009; Singh et al., 2012). Due to these phytopathogens, farmers, decision-makers, researchers, and consumers face huge challenges (Fletcher et al., 2006). According to Dean et al analysis of the top 10 plant fungal infections, farmers face a variety of challenging issues when producing plants (2012). Destructive diseases are caused worldwide by plant fungal pathogens such Magnaporthe oryzae and Colletotrichum spp. Blumeria graminis decreases crop amount while Fusarium graminearum lowers crop quality. One disease, such as Mycosphaerella graminicola, can mutate and infect different plant species in a same field, diminishing the ability of plants to fight off the infection. Similar to F. oxysporum, which possesses over 70 formae speciales, it can be challenging to pinpoint the important pathogen. A single species can spread disease to a wide range of plant species; for instance, F. oxysporum and Botrytis cinerea each have 200 plant hosts. A single pathogen, such as M. oryzae, can cause a significant reduction in grain output. Infections can fully shut down a plant's immune system when they coexist with other pathogens, such as F. graminearum and other Fusarium species. Additionally, pathogens like M. graminicola have a longer than 7-day symptomless colonisation period. Because of this, it is challenging to tell if the plant is diseased. Because some infections, like Ustilago maydis, can complete their life cycle in as little as two weeks, they can be extremely harmful. It is challenging to even produce other crops in crop rotation as a result of pathogens like Puccinia spp. Some pathogens have the ability to infect new plant species, afflict several plant species, or inflict disease on related plant species. Numerous weak infections can wreak havoc and start epidemics in closely related species across continents (Burdon and Thrall, 2009). In the pre- and post-harvest processes, plant fungi infections cause annual economic losses of more than \$200 billion USD (Horbach et al., 2011). (Gonzalez-Fernandez et al., 2010). A billion dollars in yield and quality losses were attributed to the emergence of wheat and barley scab (cause: F. graminearum) in North America in 1993 alone (Mullins and Kang, 2001). It is obvious that fungi pose a wide range of issues and have the ability to have a significant impact on plant growth.

B. Plant Fungi That Cause Post-Harvest Issues And Food Deterioration

The customer may be exposed to post-harvest illnesses and food spoilage brought on by plant fungi at any time during the processing process, including during harvesting, handling, storage, packaging, and transportation (Agrios, 2005). Fresh fruit and vegetable rot in postharvest procedures is mostly caused by fungi (Gatto et al., 2011). The majority of postharvest infections are caused by more than 100 species of fungi, and they can reduce crop output by 10 to 30 percent (Tripathi and Dubey, 2004). (Agrios, 2005). The loss of perishable goods might reach 50% in impoverished nations and tropical areas (Tripathi and Dubey, 2004). Due to microbial deterioration, certain plant fungal diseases generate considerable losses during storage. Fungi include *Penicillium italicum* and *P. digitatum* (green rot of citrus), *Penicillium expansum* (blue rot of apples and pears), *Penicillium glabrum* (onion), and *Penicillium funiculosum* (onion), as well as *Botrytis cinerea* (fruits such as raspberries, strawberries, grapes, kiwi fruit, pears, peaches, plums, and cherries; vegetables such as (Moss, 2008). *Fusarium*, *Geotrichum*, and *Aspergillus* are a few fungal species that cause fruit and vegetable deterioration, leading to considerable financial losses and unpleasant features in plant-based products (Agrios, 2005). Fruits that have been preserved can be completely destroyed by *colletotrichum* infections (Dean et al., 2012).

C. Issues Brought On By Mycotoxins

Molds create mycotoxins, which are low-molecular-weight substances. These secondary fungal metabolites have no impact on the formation or growth of the fungus and are poisonous to vertebrates at very low quantities (Hussein and Brasel, 2001). Mycotoxins are present in a wide variety of foods and are significant because they can result in both human and animal disorders like cancer and dermatitis (Agrios, 2005; Kumar et al., 2008). Consumption of tainted plant foods or animal feed containing mycotoxins can also result in a variety of metabolic issues such liver function decline, interference with protein synthesis, or other conditions like skin sensitivity, necrosis, or severe immunodeficiency (Sweeney and Dobson, 1998). Up to 25% of global crops are infected with mycotoxins every year, which has a serious impact on food security and the economy by causing the loss of 1 billion tonnes of food and food products (Matny, 2015). *Aspergillus*, *Fusarium*, and *Penicillium* species are frequently responsible for the mycotoxin-related illnesses (Agrios, 2005). Aside from having detrimental impacts on human and animal health, toxins such aflatoxins, ochratoxins, trichothecenes, zearalenone, fumonisins, tremorgenic toxins, and ergot alkaloids also have a negative economic and agricultural impact (Hussein and Brasel, 2001). Therefore, plant fungal diseases play a very detrimental impact in the efforts to feed the world's ever-increasing population. Plant fungal infections and the toxins they create seriously endanger agricultural production by reducing the quality and quantity of plant commodities, which causes financial losses both before and after harvesting. Various management and control methods are utilised to try and reduce the harm that fungi can do in agricultural settings.

D. Preventing Plant Pathogens

Many people around the world would go hungry and suffer from some of the terrible illnesses and outbreaks that occasionally occur if there were no control over or induced plant defences. Various agrochemical agents have been developed and utilised to combat the recurring attack of plant fungal infections on agricultural crops. Some of these agrochemicals are hazardous to people, so there must be a waiting period before harvesting after the final dose. Numerous of these fungicides also harm plant pollinators, insects, and soil-dwelling creatures. Farmers are paying increasing attention to using natural ingredients in the

manufacturing of organic food, which is strongly backed by environmentalists and certain customers.

E. Building Up Fungal Resistance

Utilizing plant defence molecules in agricultural production to promote resistance to invasive fungal infections is the foundation of natural resistance (Nega, 2014). When different crops are plagued by diseases, salicylic acid and its analogues are utilised to create systemic acquired resistance. According to research, 30 g of benzo(1,2,3)thiadiazole-7-carbothioic acid S-methyl ester (BTH) can shield wheat crops from *Puccinia recondita* and *Septoria* species for an entire season (Reignault and Walters, 2007). Jasmonic acid and its derivatives can increase crop resistance and the synthesis of chemicals that are beneficial to health (Wasternack, 2014). Methyl jasmonates prevent degradation of "Marsh Seedless" grapefruit by *P. digitatum* and postharvest infections of strawberries by *B. cinerea* in agricultural settings (Tripathi and Dubey, 2004). Other natural substances that promote resistance against fungi include chitosan, -aminobutyric acid, glucosinolates, propolis, fusapyrone, deoxyfusapyrone, ethephon, microbial products, and plant extracts. These goods are utilised in agricultural settings all over the world to improve quality and yields (Tripathi and Dubey, 2004; Thakur and Sohal, 2013).

F. Fungicides

Synthetic compounds have offered much-needed relief in the management of plant fungal disease in agricultural output since the initial use of fungicides in the 1800s. Over time, the use of diverse synthetic compounds in agricultural production has reduced the impact of numerous plant fungi diseases, boosted plant crop yield, and generated profits (Gianessi and Reigner, 2006). Farmers, however, have struggled with the growth of fungicide resistance since the 1970s (Ishii, 2006; Possiede et al., 2009). This causes financial losses to occur. Fungicide sales were roughly 5.9 billion US dollars worldwide in 1996. (Martinez, 2012). The USA spends more than 600 million dollars on synthetic chemicals every year (Gonzalez-Fernandez et al., 2010). Japan had the largest fungicide market in the world in 2002, with sales of 818 million US dollars (Ishii, 2006). Fungicides frequently cause environmental imbalances by being poisonous to non-target animals like earthworms, bacteria, and people (genotoxicity) (Nega, 2014, Patel et al., 2014). Numerous of these compounds decay slowly and are challenging to eliminate. These might contaminate rivers and waterways as well (Stamatis et al., 2010).

G. Biological Defence

Numerous scientists are looking for alternatives to synthetic chemicals that are biological in order to address the issues with such substances (Martinez, 2012; Nega, 2014). Chemicals generated from microbial, botanical, and animal sources are classified as biological pesticides. At the moment, there are hundreds of products using more than 245 registered biopesticide-active chemicals in the USA. These make about 20% of all pesticide active chemicals that are registered in the nation (Yoon et al., 2013). Based on the fact that bacteria are antagonistic to the fungi that cause fungal illnesses in plants, they may be used in their treatment. The findings of investigations using microbial antagonists against plant fungal infections in the lab and in the field are encouraging (Sharma et al., 2009). Numerous fungal and bacterial antagonistic commercial products, such as GiloGard (*Trichoderma harzianum* - several soilborne illnesses), F-Stop (*Gliocladium virens* - seedling diseases of ornamentals and bedding plants), *Agrobacterium radiobacter* K-84, Gallex, or Galltrol (*T. harzianum*/*T. polysporum* - to prevent wood deterioration), Problems related to plant fungal infections are successfully treated with Dagger G (*Pseudomonas fluorescens* - *Rhizoctonia*

and Pythium damping-off of cotton) and Kodiak (*Bacillus subtilis* - seed illnesses) around the world (Agrios, 2005).

H. The Use Of Plant Extracts

Numerous studies on the *in vitro* antifungal activity of plant extracts have been reported. The reason why many antifungal chemicals in plant extracts are very non-polar and do not disperse well in the aqueous agar matrix is why many authors, unfortunately, have employed techniques like agar diffusion experiments that do not perform well with plant extracts. Because so many factors affect the findings of agar diffusion, it is also very challenging to compare data between various laboratories. Different strategies for defending people or animals against fungus have been covered by Eloff and McGaw (2006) and McGaw and Eloff (2010). To determine the antibacterial activity of plant extracts, a serial dilution technique using tetrazolium violet as an indicator of growth was created and is now widely used (Eloff, 1998a). This approach has been improved and is excellent for fungi (Masoko et al., 2005). This technique allows for the calculation of the extracts' minimum inhibitory concentration (MIC). Only MICs of 0.1 mg/ml and lower should be thought of as having significant activity, according to several authors (Eloff, 2004; Rios and Recio, 2005; Cos et al., 2006). Using bioautography, it was also possible to determine how many antifungal chemicals were present in plant extracts (Masoko and Eloff, 2005). Acetone was typically the best extract when multiple extractants were employed (Eloff, 1988b). Water extracts often exhibited relatively little antibacterial action (Kotze and Eloff, 2002). Traditional leads might not be very helpful because water is the major extractant available to traditional healers. As a result, acetone tree leaf extracts were randomly tested for efficacy against nosocomial bacteria and fungus. Only a few extracts, many of which had MICs of 0.02 mg/ml and lower, had a MIC higher than 2,5 mg/ml when the antibacterial activity of 717 crude extracts of 537 tree species was assessed against four bacteria and two fungi (Pauw and Eloff, 2014). The antifungal properties of numerous plant species have been researched (Eloff and McGaw, 2014; Raut and Karuppayil, 2014). In storage circumstances, many essential oils prevent post-harvest fungal infections and extend the shelf life of many crops (Tripathi and Dubey, 2004). Additionally, a variety of fungi species formation of mycotoxin is inhibited by essential oils (Sivakumar and Bautista-Banos, 2014).

It is obvious that fungi provide significant challenges to the plant production sector and that poor management might have detrimental effects on the ability to produce food. Fungi can have an impact on the production of medicinal plants as well as their safety after harvesting in the same ways that phytopathogens do on the food and floricultural industries. In order to deal with the advent or outbreaks of plant fungal diseases, the existing control strategies are insufficient (Ishii, 2006; Possiede et al., 2009). Consequently, further research is needed, particularly using plant-based goods, to produce affordable, safe, and efficient biological products (Martinez, 2012). Fighting plant fungal diseases may be made easier by control using plant-based products (Tripathi and Dubey, 2004). Despite extensive study on testing plants for their antifungal activity against phytopathogens, only a small number of secondary metabolites have been identified (Cowan, 1999). Finding novel plant-based antifungal chemicals is one strategy, but another option is to use a complex plant extract. The benefit of the latter strategy is that, if the various antifungal components in an extract target various receptors, resistance may be less likely to develop. However, there is a drawback in terms of maintaining high quality control and activity variation due to genetic or environmental factors when compared to employing a single chemical product.

CONCLUSION

Research on plants that can defend plants against fungi is a fruitful and significant topic of study, and there have been many publications on the use of plant products in human or animal medicine. Although an essential field, employing plant products to give organically grown plants or medicine won't soon replace chemical antifungals. Aspects of quality control raise the cost of plant production, harvesting, and extraction. However, using plant-based treatments to treat plants during the period before harvesting could be crucial to agriculture in order to allow the concentration of chemical control agents to fall to acceptable levels. Even if there have been several publications on the use of plant products in human or animal medicine, research on plants that can defend plants against fungi or against livestock could be a useful field of study to improve the quality of life for people living in rural areas.

REFERENCES

- Al-Awkally, Noor Alhooda Milood, Ibrahim, Hamza Khalifa, & Samad, Abdul. (2022). Antipsychotic Combinations for Psychiatric Disorders. *BULLET: Jurnal Multidisiplin Ilmu*, 1(01), 49–50.
- Agrios, G.N. (2009). Plant pathogens and disease: general introduction. Elsevier Inc., University of Florida, Gainesville, FL, USA.
- Agrios, G.N. (2005). Plant pathology. Fifth edition. Elsevier Acad Press, Amsterdam.
- Alexandratos, N. and Bruinsma, J. (2012). World agriculture towards 2030/2050: the 2012 revision. ESA Working Paper Rome, FAO.
- Al-Awkally, Noor-Alhooda Milood, Hamza Khalifa Ibrahim, and Abdul Samad. "Antipsychotic Combinations for Psychiatric Disorders." *BULLET: Jurnal Multidisiplin Ilmu* 1.01 (2022): 49-50
- Anderson, P.K., Cunningham, A.A., Patel, N.G., Morales, F.J., Epstein, P.R. and Daszak, P. (2004). Emerging infectious diseases of plants: pathogen pollution, climate change and agrotechnology drivers. *Trends Ecol. Evol.*, 19:10, 535 – 544.
- Burdon, J.J. and Thrall, P.H. (2009). Plant pathogens and disease: newly emerging diseases. Elsevier Inc.
- Chitarra, G.S., Abee, T., Rombouts, F.M., Posthumus, M.A. and Dijksterhuis, J. (2004). Germination of *Penicillium paneum* conidia is regulated by 1-octen-3-ol, a volatile self-inhibitor. *Appl. Environ. Microbiol.*, 70: 5, 2823 – 2829.
- Cos, P., Vlietinck, A.J., Vanden Berghe, D. and Maes, L. (2006). Anti-infective potential of natural products: how to develop a stronger in vitro 'proof-of-concept'. *J. Ethnopharmacol.*, 106: 290 – 302.
- Cowan, M.M. (1999). Plant products as antimicrobial agents. *Clin. Microbiol. Rev.*, 12: 564 – 582.
- Dean, R., Van Kan, J.A.L., Pretorius, Z.A., Kosack, K.E.H., Di Pietro, A., Spanu, P.D., Rudd, J.J., Dickman, M., Kahmann, R., Ellis, J. and Forster, G.D. (2012). The Top 10 fungal pathogens in molecular plant pathology. *Mol. Plant Pathol.*, 13: 4, 414 – 430.
- De Silva, N.I., Lumyong, S., Hyde, K.D., Bulgakov, T., Phillips, A.J.L. and Yan, J.Y. (2016). Mycosphere essays 9: defining biotrophs and hemibiotrophs. *Mycosphere*, 7: 5, 545 – 559.
- El Hussein, A.A., Alhasan, R.E.M., Abdelwahab, S.A. and El Siddig, M.A. (2014). Isolation and identification of *Streptomyces rochei* strain active against Phytopathogenic Fungi. *Br. Microbiol. Res. J.*, 4: 10 1057 – 1068.

- Ellis, S.D., Boehm, M.J. and Mitchell, T.K. (2008). Fungal and fungal-like diseases of plants. Fact Sheet (PP401.07) Agriculture and natural resources, the Ohio State University.
- Eloff, J.N. (1998a). Which extractant should be used for the screening and isolation of antimicrobial components from plants? *J. Ethnopharmacol.*, 60: 1 – 8.
- Eloff, J.N. (1998b). A sensitive and quick microplate method to determine the minimal inhibitory concentration of plant extracts for bacteria. *Planta Med.*, 64: 711 – 714.
- Eloff, J.N. (2004). Quantifying the bioactivity of plant extracts during screening and bioassay-guided fractionation. *Phytomedicine*, 11: 370-371.
- Eloff, J.N. and McGaw, L.J. (2006). Plant extracts used to manage bacterial, fungal and parasitic infections in southern Africa. Ahmad I (Edit.) *Modern phytomedicine: turning medicinal plants into drugs* Wiley-VCH, Germany, 97 – 121.
- Eloff, J.N., Angeh, I. and McGaw, L.J. (2007). A potentised leaf extract of *Melianthus comosus* has higher activity against six commercial products used against plant fungal pathogens. *S. Afr. J. Bot.*, 73: 286.
- Eloff, J.N. and McGaw, L.J. (2014). Using African plant biodiversity to combat microbial infections. Page 163 – 173 in Gurib-Fakim A (Ed) *Novel Plant Bioresources: Applications in Food Medicine and Cosmetics*. John Wiley DOI: 10.1002/9781118460566.ch12.
- Fletcher, J., Bender, C., Budowle, B., Cobb, W.T., Gold, S.E., Ishimaru, C.A., Luster, D., Melcher, U., Murch, R., Scherm, H., Seem, R.C., Sherwood, J.L., Sobral, B.W. and Tolin, S.A. (2006). Plant pathogen forensics: capabilities, needs, and recommendations. *Microbiol. Mol. Biol. Rev.*, 70: 450 – 471.
- Gatto, M.A., Ippolito, A., Linsalata, V., Cascarano, N.A., Nigro, F., Vanadia, S. and Di Venere, D. (2011). Activity of extracts from wild edible herbs against postharvest fungal diseases of fruit and vegetables. *Postharvest Biol. Technol.*, 61: 72 – 82.
- Gianessi, L. and Reigner, N. (2006). The importance of fungicides in U.S. crop production. *Outlook. Pest Manag.* 10:209 – 213.
- Gonzalez-Fernandez, R., Prats, E. and Jorin-Novo, J.V. (2010). Proteomics of plant pathogenic fungi. *J. Biomed. Biotechnol.*, 2010: 1 – 36.
- Gould, A.B. (2009). *Fungi: plant pathogenic*. Elsevier Inc., 457 – 477.
- Gurib-Fakim, A. (2006). Medicinal plants: Traditions of yesterday and drugs of tomorrow. *Mol. Aspects Med.*, 27: 1 –33.
- Horbach, R., Navarro-Quesada, A.R., Knogge, W. and Deising, H.B. (2011). When and how to kill a plant cell: infection strategies of plant pathogenic fungi. *J. Plant Physiol.*, 168: 51 – 62.
- Hussein, S.H. and Brasel, J.M. (2001). Toxicity, metabolism, and impact of mycotoxins on humans and animals. *Toxicol.*, 167: 101– 134.
- Ishii, H. (2006). Impact of fungicide resistance in plant pathogens on crop disease control and agricultural environment. *Jpn. Agric. Res. Q.*, 40: 3, 205 – 211.
- Ibrahim, H. K., Al-Awkally, N. A. M., Samad, A., Zaib, W., & Hamza, M. (2022). Covid-19 Pandemic and Its Impact on Psychological Distress, Malignancy and Chronic Diseases: A Scoping Review. *Eduvest-Journal Of Universal Studies*,

- 2(5), 1017-1021.
- Knogge, W. (1996). Fungal infection of plants. *Plant Cell*, 8: 1711 – 1722.
- Knogge, W. (1998). Fungal pathogenicity. *Curr. Opin. Plant Biol.*, 1: 324 – 328.
- Kotze, M. and Eloff, J.N. (2002). Extraction of antibacterial compounds from *Combretum microphyllum*(Combretaceae). *S. Afr. J. Bot.*, 68: 62 – 67.
- Kumar, V., Basu, M.S. and Rajendran, T.P. (2008). Mycotoxin research and mycoflora in some commercially important agricultural commodities. *Crop Prot.*, 27: 891 – 905.
- Mahlo, S.M. and Eloff, J.N. (2014). Acetone leaf extracts of *Breonadia salicina* (Rubiaceae) and ursolic acid protects oranges against infection by *Penicillium* species. *S. Afr. J. Bot.*, 93: 48 – 53.
- Mahlo, S.M., McGaw, L.J. and Eloff, J.N. (2010). Some tree leaf extracts have good activity against plant fungal pathogens. *Crop Prot.*, 29: 1529 – 1533.
- Mahlo, S.M., McGaw, L.J. and Eloff, J.N. (2013). Antifungal activity and cytotoxicity of isolated compounds from leaves of *Breonadia salicina*. *J. Ethnopharmacol.*, 148: 909 – 913.
- Martínez, J.A. (2012). Natural fungicides obtained from plants, fungicides for plant and animal diseases, Dr. Dharumadurai Dhanasekaran (Ed.), ISBN: 978-953-307-804-5, InTech, DOI: 10.5772/26336.
- Masoko, P. and Eloff, J.N. (2005). The diversity of antifungal compounds of six South African *Terminalia* species (Combretaceae) determined by bioautography. *Afr. J. Biotechnol.*, 4: 1425 – 1431.
- Masoko, P., Picard, J. and Eloff, J.N. (2005) Antifungal activities of six South African *Terminalia* species (Combretaceae). *J. Ethnopharmacol.*, 99: 301 – 308.
- Matny, O.N. (2015). *Fusarium* head blight and crown rot on wheat & barley: losses and health risks. *Adv. Plants Agr.Res.*, 2: 2 – 7.
- McGaw, L.J. and Eloff, J.N. (2010). Methods for evaluating efficacy of ethnoveterinary medicinal plants, page 1 – 24. In Katerere DR and Luseba D (Eds) *Ethnoveterinary Botanical Medicine: Herbal medicines for Animal Health* CRC Press London.
- Mdee, L.K., Masoko, P. and Eloff, J.N. (2009). The activity of extracts of seven common invasive plant species on fungal phytopathogens. *S. Afr. J. Bot.*, 75: 375 – 379.
- Mohammed, A. A., Samad, A., & Omar, O. A. (2022). *Escherichia coli* spp, *Staph albus* and *Klebseilla* spp were affected by some Antibiotics for Urinary Tract Infections in Bani Waleed City. *Brilliance: Research of Artificial Intelligence*, 2(2), 66-70.
- Moss, M.O. (2008). Fungi, quality and safety issues in fresh fruits and vegetables. *J. of Appl. Microbiol.*, 104: 1239 –1243.
- Mullins, E.D. and Kang, S. (2001). Transformation: A tool for studying fungal pathogens of plants. *Cell. Mol. Life Sci.*, 58: 2043 – 2052.
- Muthanna, F. M., & Samad, A. (2022). Covid-19 Pandemic (Incidence, Risk factors and Treatment). *BULLET: Jurnal Multidisiplin Ilmu*, 1(01), 46-48.
- Muthanna, F. M., Samad, A., Ibrahim, H. K., Al-Awkally, N. A. M., & Sabir, S. (2022). Cancer related anaemia (CRA): An overview of approach and treatment. *International Journal of Health Sciences*, 6, 2552-2558.

- Nega, A. (2014). Review on concepts in biological control of plant pathogens. *J. Biol. Agric. and Healthc.*, 4: 27, 33 –35.
- Okwute, S.K. (2012). Plants as potential sources of pesticidal agents: a review, pesticides - advances in chemical and botanical pesticides, Dr. R.P. Soundararajan (Ed.), InTech, DOI: 10.5772/46225.
- Osharov, N. and May, G. (2001). The molecular mechanisms of conidial germination. *FEMS Microbiol. Lett.*, 199: 153– 160.
- Patel, N., Desai, P., Patel, N., Jha, A. and Gautam, H.K. (2014). Agronanotechnology for plant fungal disease management: a review. *Int. J. Curr. Microbiol. App. Sci.*, 3: 10, 71 – 84.
- Pauw, E. and Eloff, J.N. (2014). Which tree orders in southern Africa have the highest antimicrobial activity and selectivity against bacterial and fungal pathogens of animals? *BMC Complement. Altern. Med.*, 14:317 doi:10.1186/1472-6882-14-317 (8 pages).
- Podila, G.K., Rogers, L.M. and Kolsttukudy, P.E. (1993). Chemical signals from avocado surface wax trigger germination and appressorium formation in *Colletotrichum gloeosporioides*. *Plant Physiol.*, 103: 267 – 272.
- Possiede, Y.M., Gabardo, J., Kava-Cordeiro, V., Galli-Terasawa, L.V., Azevedo, J.L. and Glienke, C. (2009). Fungicide resistance and genetic variability in plant pathogenic strains of *Guignardia citricarpa*. *Braz. J. Microbiol.*, 40: 308 – 313.
- Raut, J.S. and Karuppayil, S.M. (2014). A status review on the medicinal properties of essential oils. *Ind. Crops Prod.*, 62: 250 – 264.
- Reignault, P. and Walters, D. (2007). Topical induction of inducers for disease control. D. Walters, A. Newton, G. Lyon (Eds.), *Induced resistance for plant disease control: a sustainable approach to crop protection*, Blackwell Publishing, Oxford.
- Ribera, A.E. and Zuniga, G. (2012). Induced plant secondary metabolites for phytopathogenic fungi control: a review. *J. Soil Sci. Plant Nutr.*, 12: 4, 893 – 911.
- Rios, J.L. and Recio, M.C. (2005) Medicinal plants and antimicrobial activity. *J Ethnopharmacol.*, 100: 80 – 84.
- Rossmann, A.Y. (2009). The impact of invasive fungi on agricultural ecosystems in the United States. *Biol. Invasions.*, 11: 97 – 107.55. Schafer, W. (1994). Molecular mechanisms of fungal pathogenicity to plants. *Annu. Rev. Phytopathol.*, 32: 461 – 477. Sharma, R.R., Singh, D. and Singh, R. (2009). Biological control of postharvest diseases on fruits and vegetables by microbial antagonists: a review. *Biol. Control.*, 50: 205 – 221.
- Samad, A., Ahmad, H., Hamza, M., Muazzam, A., Ahmer, A., Tariq, S & Muthanna, F. M. (2022). Overview of Avian Corona virus, its prevention and control Measures. *BULLET: Jurnal Multidisiplin Ilmu*, 1(01), 39-45.
- Samad, A. , Hamza , M., Muazzam, A. , Ahmad, H. , Ahmer, A. , Tariq, S. , Khera, H. U. R. A. , Mehtab, U. , Shahid, M. J. , Akram, W. , Kaleem, M. Z. , Ahmad, S. , Abdullah, A. , & Ahmad, S. . (2022). Policy of control and prevention of infectious bursal disease at poultry farm. *African Journal of Biological, Chemical and Physical Sciences*, 1(1), 1-7.
- Samad A, Abbas A, Mehtab U, Ur Rehman Ali Khera H, Rehman A and Hamza M . Infectious Bronchitis Disease in Poultry its Diagnosis, Prevention and Control

- Strategies. *Ann Agric Crop Sci.* 2021; 6(7): 1100.
- Samad, A., Hamza, M., Muazzam, A., Ahmer, A., Tariq, S., Shahid, M. J., ... & Din, F. U. (2022). Overview of Bacterial Diseases in Poultry and policies to control disease and antibiotic resistance. *BULLET: Jurnal Multidisiplin Ilmu*, 1(01), 19-25.
- Samad, A., Hamza, M., Muazzam, A., Ahmer, A., Tariq, S., Javaid, A., ... & Ahmad, S. (2022). Newcastle Disease In Poultry, Its Diagnosis, Prevention And Control Strategies. *BULLET: Jurnal Multidisiplin Ilmu*, 1(01), 1-5.
- Schafer, W. (1994). Molecular mechanisms of fungal pathogenicity to plants. *Annu. Rev. Phytopathol.*, 32: 461 – 477.
- Sharma, R.R., Singh, D. and Singh, R. (2009). Biological control of postharvest diseases on fruits and vegetables by microbial antagonists: a review. *Biol. Control.*, 50: 205 – 221.
- Shuping DSS (2016). Development of an antifungal product from *Melianthus comosus* (Melianthaceae) that can be used to control plant fungal pathogens. PhD thesis, University of Pretoria, Pretoria, South Africa.
- Singh, D., Jackon, G., Hunter, D., Fullerton, R., Lebot, V., Taylor, M., Iosefa, T., Okpul, T. and Tyson, J. (2012). Taro leaf blight – a threat to food security. *Agric.*, 2: 182 – 203.
- Sivakumar, D. and Bautista-Banos, S. (2014). A review on the use of essential oils for postharvest decay control and maintenance of fruit quality during storage. *Crop Prot.*, 64: 27 – 37.
- Stamatis, N., Helac, D. and Konstantinou, I. (2010). Occurrence and removal of fungicides in municipal sewage treatment plant. *J. Hazard. Materi.*, 175: 829 – 835.
- Stergiopoulos, I. and de Wit, P.J.G.M. (2009). Fungal Effector Proteins. *Annu. Rev. Phytopathol.*, 47: 233 – 263.
- Strobel, G. and Daisy, B. (2003). Bioprospecting for microbial endophytes and their natural products. *Microbiol. Mol.Biol. Rev.*, 67
- Sweeney, M.J. and Dobson, A.D.W. 1998. Mycotoxin production by *Aspergillus*, *Fusarium* and *Penicillium* species. *Int. J. Food Microbiol.*, 43: 141 – 158.
- Thakur, M. and Sohal, B.S. (2013). Role of elicitors in inducing resistance in plants against pathogen infection: A review. *ISRN Biochem.*, 1: 1 – 10.
- Thomma, B.P.H.J., Nurnberger, T. and Joosten, M.H.A.J. (2011). Of PAMPs and effectors: The blurred PTI-ETI dichotomy. *Plant Cell*, 23: 1, 4 – 15.
- Tripathi, P. and Dubey, N.K. (2004). Exploitation of natural products as alternative strategy to control post-harvest fungal rotting of fruits and vegetables. *Postharvest Biol. Technol.*, 32: 235 – 245.
- Wasternack, C. (2014). Action of jasmonates in plant stress responses and development-applied aspects. *Biotechnol.Adv.*, 32: 31 – 39.68.
- Wen, L. (2013). Cell death in plant immune response to necrotrophs. *J. Plant Biochem. Physiol.*, 1: 1 – 3.
- Yoon, M-Y., Cha, B. and Kim, J-C. (2013). Recent trends in studies on botanical fungicides in agriculture. *Plant Pathol. J.*, 29: 1, 1 – 9.
- Zvereva, A.S. and Pooggin M.M. (2012). Silencing and innate immunity in plant defense against viral and non-viral pathogens. *Virus.*, 4: 2578 – 2597

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